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EFFECT OF FOLIAR APPLICATION OF ZNO NANOPARTICLES ON SEED QUALITY OF COWPEA (*Vigna unguiculata* L. Walp)

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ABSTRACT

The zinc oxide nanoparticles (ZnO NPs) were synthesized through biological method using leaf extract of *Azadirachta indica*. The efficacy of green synthesized ZnO NPs was attributed on seed quality of cowpea. The present investigation was carried out during *kharif* 2023 and 2024 using two varieties of cowpea V₁: Phule Sonali and V₂: Phule Rakhumai to determine effect of the foliar application of green synthesized zinc oxide nanoparticles (ZnO NPs) which was undertaken at 30, 45 and 60 days after sowing at different concentrations as a treatments viz. T₀: Untreated Control, T₁: Zinc oxide nanoparticle (ZnO NPs) @ 50 ppm, T₂: Zinc oxide nanoparticle (ZnO NPs) @ 75 ppm, T₃: Zinc oxide nanoparticle (ZnO NPs) @ 100 ppm, T₄: Zinc oxide nanoparticle (ZnO NPs) @ 125 ppm and T₅: Zinc oxide nanoparticle (ZnO NPs) @ 150 ppm. The foliar application of green synthesized ZnO NPs @ 125 ppm (T₄) had higher 100 seed weight, germination percentage, seedling length, seedling dry weight, vigour index I and vigour index II with lower electrical conductivity and seed mycoflora percentage followed by ZnO NPs @ 150 ppm (T₅) and which was superior over control (T₀).

Keywords: Green synthesis, seed quality, zinc oxide nanoparticles, cowpea, seed quality.

Introduction

Cowpea (*Vigna unguiculata* L. Walp.) is a diploid species (2n=2x=22). It is an annual legume belonging to family Fabaceae (Maréchal *et al.*, 1978) or Leguminosae and the subfamily Papilionaceae (Cobley and Steele, 1976). However, Sellsschop (1962) and Purseglove (1976) stated that all cultivars belong to the wild cowpea, *Vigna unguiculata* L. Walp, which is indigenous to tropical Africa (central Africa). In India zinc (Zn) is considered as fourth most important yield limiting nutrient in agricultural crops. Zinc deficiency in Indian soils is likely to increase from 49 to 63 per cent by 2025 (Arunachalam *et al.*, 2013). Zinc is required for chlorophyll production, pollen function, fertilization, germination and for biomass production (Pandey *et al.*, 2006). Sufficient

amounts of micronutrients are required for optimal cowpea growth and development. These include zinc (Zn) because its deficiency negatively affects plant reproductive functions and pollen development, reduces leaf area, induces chlorosis and abnormal growth of plant structures (Pandey *et al.*, 2006). This deficiency can also result in higher sensitivity to biotic and abiotic stress (Obata *et al.*, 1999). Most importantly, Zn agronomic management includes foliar application and soil fertilization. Zinc involved in photosynthesis, DNA replication and transcription, hydrocarbon metabolism, regulation of auxin production and cell membrane integrity (Sturikova *et al.*, 2018). Zinc is a micro nutrient involved in many aspects of plant metabolic and structural process such as photosynthesis, respiration, the production of

chlorophyll and chloroplast function (Zulfiqar *et al.*, 2020).

Zinc oxide is regarded as one of the most important nano materials among nanoparticles because of its stability, non-toxicity, low cost of manufacture, and environmental friendliness. The physiochemical properties of ZnO nanoparticles allow them to penetrate the plant tissue and interfere with several metabolic processes. According to Miralles *et al.* (2012), ZnO can pierce the seed coat and induce embryonic differentiation by activating enzymes involved in seed dormancy breakdown. Zinc oxide nanoparticles not only enhance seed germination but they also improve seed longevity (Gayathri, 2019). Zinc oxide nanoparticles are also shown to have a beneficial effect on the response of phytohormones, particularly Indole Acetic Acid (IAA), enhancing phytostimulatory effects. Zinc-rich ZnO NPs may enhance the level of IAA in roots (sprouts), hence increasing seedling development rates (Pandey *et al.*, 2010). Zinc nanoparticles (ZnO NPs) have received significant interest because of their low cost, large surface area, appearance, UV-filtering, antifungal, antibacterial and photochemical features and high catalytic activity. Recent studies have demonstrated ZnO efficacy as a fungicide due to their ability to prevent the growth of disease-causing fungi in plants. Applications of nanotechnology in improving seed germination, emergence and growth of seedlings (Zhang *et al.*, 2006) and for early pathogen detection (Alocilja and Radke, 2003) are few of the multifarious beneficial interventions in the field of agriculture.

Materials and Methods

The field experiment was laid down during *kharif*, 2023 and 2024 in Factorial Randomized Block Design (FRBD) with three replications and the experiment was conducted at the field of Botany Farm, MPKV, Rahuri. The seed of cowpea cultivars V₁: Phule Sonali and V₂: Phule Rakhumai used for this experiment. The foliar application of green synthesized zinc oxide nanoparticles (ZnO NPs) was undertaken at 30, 45 and 60 days after sowing using treatments T₀: Untreated Control, T₁: Zinc oxide nanoparticle (ZnO NPs) @ 50 ppm, T₂: Zinc oxide nanoparticle (ZnO NPs) @ 75 ppm, T₃: Zinc oxide nanoparticle (ZnO NPs) @ 100 ppm, T₄: Zinc oxide nanoparticle (ZnO NPs) @ 125 ppm and T₅: Zinc oxide nanoparticle (ZnO NPs) @ 150 ppm.

Observations recorded

100 seed weight (g)

Hundred seeds in each treatment were counted manually and the weight was measured by using

electronic weighing balance. The hundred seed weight was recorded in grams.

Electrical conductivity (dsm⁻¹)

Five (5) g of seeds were soaked in 25 ml distilled water and kept in incubator maintained at 25 ± 1° C for 24 hrs. After soaking and gently swirling the solution. The seeds were discarded and the leachate decanted and final volume made up to 25 ml by adding distilled water. This leachate was used for measuring the electrical conductivity with help of digital electrical conductivity meter and expressed in dSm⁻¹ (Presley, 1958).

Germination (%)

Four replications of 100 seed from each treatment were kept for germination at 25±1°C for 10 days in between paper (BP) method. The germination percentage was expressed on the basis of normal seedlings as described in ISTA Rules (Anon., 2013).

$$\text{Germination (\%)} = \frac{\text{Number of normal seedlings}}{\text{Total numbers of seeds}} \times 100$$

Seedling length (cm)

Ten normal seedlings in each replication were randomly selected at final count of the germination test and the length was measured from the tip of the primary root to the point of attachment of cotyledons. The mean of ten seedling length was expressed in centimetres.

Seedling dry weight (g)

Ten normal seedlings were selected for seedling length measurement after removing cotyledons (remnant seed) were kept in an oven maintained at 80 ± 1°C for 24 hours. After drying, the weight of dry seedlings was recorded and means weight was calculated and expressed in grams.

Vigour index I

At the end of germination test ten normal seedlings from each replication were selected for calculation of vigour index by adopting the formula suggested by (Abdul- Baki and Anderson, 1973) and expressed in whole number as given below,

$$\text{Vigour index I} = \text{Germination (\%)} \times \text{Mean seedling length (cm)}$$

Vigour index II

Vigour index II was computed by multiplying standard germination percentage and mean seedling dry weight by adopting the formula suggested by Abdul Baki and Anderson (1973) and expressed in whole number as given below,

Vigour index II = Germination (%) × Mean seedling dry weight (g)

Seed mycoflora (%)

Standard blotter paper method recommended by International Seed Testing Association (Anonymous, 1966 and Agarwal, 1976) were used to detect the seed mycoflora. Three discs of blotter paper was moistened with sterilized water; excess of water removed and placed it at the bottom of each petri-plate and labelled for each treatment. The 10 seeds in three replications of each treatment were placed at equidistance on blotter paper in each petri-plate under aseptic condition in the laminar air flow cabinet, after which the petri dishes were kept in an incubator at $20 \pm 2^{\circ}\text{C}$ for 7 days with a cycle of 12 hours light and 12 hours darkness. On the 8th day the seeds were examined for the seed mycoflora under stereoscopic binocular microscope and the observations of the fungi identified on the basis of sporulation and their fruiting structures were recorded. The percent incidence of fungi in seeds was expressed using the formula given below.

$$\text{Seed mycoflora (\%)} = \frac{\text{Number of seeds infected}}{\text{Total number of seeds incubated}} \times 100$$

Results and Discussion

100 seed weight (g)

From analysis confirmed the superiority of V_1 (Phule Sonali), which recorded a mean 100-seed weight of 10.61 g, as compared to 7.52 g in V_2 (Phule Rakhumai). Among the treatments, T_4 (ZnO NPs @ 125 ppm) recorded the highest 100-seed weight with a pooled mean of 9.85 g which was followed by T_5 (ZnO NPs @ 150 ppm), with a pooled mean of 9.55 g which was significantly superior over the control (T_0). The interaction between varieties and treatments on 100-seed weight was non-significant during both the years on pooled analysis, indicating a similar response of both varieties to the applied treatments. The highest 100-seed weight was observed under V_1T_4 (Phule Sonali + ZnO NPs @ 125 ppm) (Table 1).

Similar results were reported by Nalini *et al.* (2013) observed that foliar application of zinc at 0.1 per cent significantly improved seed size and seed weight in zinc-deficient black gram. Laware and Raskar (2014) reported in onion, Amin Farnia and Saeed Khodabandehloo (2015) found in maize. Likewise, Valadkhan *et al.* (2015) reported in chickpea. Hanumanthappa *et al.* (2019) observed higher test weight in groundnut with foliar application of nano-zinc. Dobariya and Patel (2021) reported in cowpea, Further, Tondey *et al.* (2022) in maize. Wankhade *et al.* (2025) also resulted that foliar

application of ZnO NPs @ 150 ppm enhanced 1000 seed weight in onion.

Electrical Conductivity (dS m^{-1})

Electrical conductivity differed significantly between the varieties during both the years on pooled analysis. Variety V_2 (Phule Rakhumai) which recorded significantly lower values with a pooled mean of 0.244 dS m^{-1} compared to variety V_1 (Phule Sonali) which recorded higher values with a pooled mean of 0.278 dS m^{-1} . Among the treatments T_4 (ZnO NPs @ 125 ppm) recorded the lowest values with pooled mean of 0.248 dS m^{-1} , followed by T_5 (ZnO NPs @ 150 ppm) and T_3 (ZnO NPs @ 100 ppm) were found statistically at par with T_4 and significantly superior over control treatment (T_0). The interaction between varieties and treatments was non-significant during both the years on pooled analysis. Lower values were observed under the combination V_2T_4 (Phule Rakhumai + ZnO NPs @ 125 ppm). The results are in accordance with findings of Kumar *et al.* (2023) in chickpea. They reported that dry dressing with nano ZnO at 250 mg kg^{-1} significantly lower the electrical conductivity of seed leachates. The reduced electrical conductivity observed under nano ZnO treatment was attributed to improved membrane integrity which resulted in higher seed quality (Table 2).

Seed Germination (%)

Among the varieties V_2 (Phule Rakhumai) recorded significantly higher germination with a pooled mean of 95.56 % compared to V_1 (Phule Sonali) with pooled mean of 94.08 %. This indicates a better germination potential in Phule Rakhumai compared to Phule Sonali. The highest germination percentage was recorded under treatment T_4 - ZnO NPs @ 125 ppm with pooled mean of 97.50% which was significantly superior to control and followed by T_5 - ZnO NPs @ 150 ppm with pooled mean of 96.17%. The interaction effect between varieties and treatments was found non-significant during both years as well as in pooled data. The maximum germination was observed in V_2T_4 (Phule Rakhumai + ZnO NPs @ 125 ppm) (Table 3).

The findings are in close conformity with Pandey *et al.* (2010) reported that ZnO nanoparticles enhanced seed germination in *Cicer arietinum*. Similarly, Prasad *et al.* (2012) observed in peanut. Sedghi *et al.* (2013), Gokak and Taranath (2015), Maithreyee and Gowda (2015) and Snehalbhai (2016) who documented improved germination following nano zinc treatments across the different crops such as soybean, okra, maize and groundnut respectively. Rai and Jajoo (2021) observed increased amylase activity and water uptake

in wheat seeds primed with ZnO nanoparticles. Further support is provided by Kumar *et al.* (2023) in chickpea.

Seedling Length (cm)

Among the varieties, V₂ (Phule Rakhumai) recorded significantly higher seedling length with a pooled mean of 26.24 cm compared to V₁ (Phule Sonali) with a pooled mean of 23.93 cm. The differences between the varieties were significant during both years on pooled analysis. The maximum seedling length was recorded under treatment T₄ (ZnO NPs @ 125 ppm) with a pooled mean of 27.40 cm which was significantly superior to control and followed by T₅ (ZnO NPs @ 150 ppm) with a pooled mean of 26.68 cm. The interaction between varieties and treatments was non-significant during both years on pooled analysis. The highest seedling length was observed in V₂T₄ (Phule Rakhumai + ZnO NPs @ 125 ppm (Table 4).

The results of present investigation, is in closely relate with the findings of Pandey *et al.* (2010) who reported in chickpea by increasing the activity of indole acetic acid (IAA). Dhoke *et al.* (2013) observed in mung bean. Shailesh *et al.* (2013) in mung bean indicating that nanoparticles facilitate improved nutrient uptake and metabolic activity during early growth stages. Patra *et al.* (2013) reported in cluster bean. Sedghi *et al.* (2013), reported in soybean. Gokak and Taranath (2015) observed in okra. Similarly, Maithreyee and Gowda (2015) in maize, Lakshmi *et al.* (2017) in mungbean, Rawat *et al.* (2018) in wheat, Tejaswini *et al.* (2019) reported in groundnut, Arshi *et al.* (2020) observed in tomato, Peiman *et al.*, (2022), in withania plants. Kumar *et al.* (2023) in chickpea reported that seed treatment with ZnO nanoparticles significantly enhanced seedling length.

Seedling Dry Weight (g)

Among the varieties, V₁ (Phule Sonali) recorded significantly higher seedling dry weight with a pooled mean of 0.064 g compared to V₂ (Phule Rakhumai) with a pooled mean of 0.049 g. Seedling dry weight was higher in seeds treated with ZnO nanoparticles compared to the control (T₀). The highest seedling dry weight was recorded in T₄ (ZnO NPs @ 125 ppm) with a pooled mean of 0.062 g followed by T₅ (ZnO NPs @ 150 ppm) with a pooled mean of 0.060 g. The interaction between varieties and treatments was non-significant during both years on pooled analysis. The highest seedling dry weight was observed in V₁T₄ (Phule Sonali + ZnO NPs @ 125 ppm) (Table 5).

The present findings are in close conformity with the results of Sedghi *et al.* (2013), who reported in soybean. Tarafdar *et al.* (2013) reported that foliar

application of nanoscale ZnO in cluster bean significantly enhanced plant biomass. Adhikari *et al.* (2015), who observed in maize treated with nano-ZnO compared to conventional zinc fertilizers. Mendez *et al.* (2016) who reported in *Capsicum annum*. Rawat *et al.* (2018) reported in wheat, Arshi *et al.* (2020) and Velasco *et al.* (2020), who reported in tomato and Peiman *et al.*, (2022) in withania plants.

Vigour Index I

Among the varieties V₂ (Phule Rakhumai) recorded significantly higher vigour index I compared to V₁ (Phule Sonali) during both years on pooled data. V₂ registered higher values with a pooled mean of 2493 than V₁ recorded with a pooled mean of 2254. The highest vigour index I was recorded under T₄ (ZnO NPs @ 125 ppm) with a pooled mean of 2672, which was significantly superior over control and followed by T₅ (ZnO NPs @ 150 ppm) with a pooled mean of 2564. The interaction between varieties and treatments was non-significant indicating a similar response pattern of both varieties to ZnO nanoparticle treatments. The highest vigour index I was recorded in V₂T₄ (Phule Rakhumai + ZnO NPs @ 125 ppm) (Table 6).

Vigour Index II

Among the varieties V₁ (Phule Sonali) recorded significantly higher vigour index II compared to V₂ (Phule Rakhumai) during both experimental years on pooled data. V₁ (Phule Sonali) recorded higher values with a pooled mean of 6.05 compared to V₂ (Phule Rakhumai) recorded values with a pooled mean of 4.67. Among the treatments the higher values was recorded under T₄ (ZnO NPs @ 125 ppm) with a pooled mean of 6.08 followed by T₅ (ZnO NPs @ 150 ppm) with a pooled mean of 5.75. The interaction between varieties and treatments was non-significant during both years on pooled analysis. The highest pooled vigour index II was recorded in V₁T₄ (Phule Sonali + ZnO NPs @ 125 ppm) (Table 7).

Similar findings were reported by Prasad *et al.* (2012), who observed in groundnut seeds. Maithreyee and Gowda (2015), who reported in maize. Snehalbhai (2016) in groundnut. Rawat *et al.* (2018), who reported in wheat seed. Rai-Kalal and Jajoo (2021) in wheat. Kumar *et al.* (2023) in chickpea.

Seed Mycoflora (%)

Among the varieties V₁ (Phule Sonali) recorded a seed mycoflora with a pooled mean of 0.56%, whereas V₂ (Phule Rakhumai) recorded lower pooled mean of 0.28%. All ZnO NPs treatments completely suppressed seed mycoflora, recording 0% infection during both

years on pooled analysis compared to the control (T_0) (pooled mean of 2.50%). In contrast, the interaction between varieties and treatments was non-significant. Similar results also reported by Kumar *et al.* (2023) highlighting the protective role of nano zinc in improving seed health and subsequent crop performance (Table 8).

Table 1: Effect of varieties, zinc oxide nanoparticle (ZnO NPs) and their interactions on 100 seed weight (g) of cowpea.

Year	2023			2024			Pooled		
Treat/Var.	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
T ₀	8.74	6.49	7.62	10.20	6.49	9.11	9.47	7.25	8.36
T ₁	9.14	6.59	7.86	10.22	6.59	9.18	9.68	7.36	8.52
T ₂	9.59	6.77	8.18	11.25	6.77	9.73	10.42	7.48	8.95
T ₃	9.77	6.87	8.32	11.79	6.87	10.02	10.78	7.56	9.17
T ₄	11.01	7.25	9.13	12.75	7.25	10.57	11.88	7.82	9.85
T ₅	10.39	7.07	8.73	12.49	7.07	10.38	11.44	7.67	9.55
Mean	9.77	6.84		11.45	8.21		10.61	7.52	
	V	T	VxT	V	T	VxT	V	T	VxT
SEm (±)	0.16	0.27	0.39	0.25	0.43	0.60	0.14	0.24	0.34
CD @ 1 %	0.63	1.10	NS	0.98	1.70	NS	0.53	0.92	NS

NS- Non Significant

Table 2: Effect of varieties, zinc oxide nanoparticle (ZnO NPs) and their interactions on Electrical Conductivity (dSm⁻¹) of cowpea.

Year	2023			2024			Pooled		
Treat/Var.	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
T ₀	0.329	0.278	0.304	0.272	0.278	0.252	0.301	0.255	0.278
T ₁	0.315	0.273	0.294	0.253	0.273	0.239	0.284	0.249	0.267
T ₂	0.309	0.275	0.292	0.249	0.275	0.236	0.279	0.249	0.264
T ₃	0.305	0.268	0.287	0.248	0.268	0.233	0.277	0.243	0.260
T ₄	0.291	0.259	0.275	0.235	0.259	0.220	0.263	0.232	0.248
T ₅	0.296	0.261	0.279	0.239	0.261	0.223	0.268	0.234	0.251
Mean	0.308	0.269		0.249	0.218		0.278	0.244	
	V	T	VxT	V	T	VxT	V	T	VxT
SEm (±)	0.003	0.004	0.006	0.003	0.004	0.006	0.002	0.003	0.004
CD @ 1 %	0.011	0.017	NS	0.011	0.017	NS	0.008	0.011	NS

NS- Non Significant

Table 3: Effect of varieties, zinc oxide nanoparticle (ZnO NPs) and their interactions on Germination (%) of cowpea.

Year	2023			2024			Pooled		
Treat/Var	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
T ₀	92.00 (73.65)	94.00 (75.85)	93.00 (74.75)	90.00 (71.62)	92.00 (73.65)	91.00 (72.64)	91.00 (72.64)	93.00 (74.75)	92.00 (73.69)
T ₁	93.00 (74.68)	95.67 (78.10)	94.33 (76.39)	92.00 (73.65)	94.00 (75.95)	93.00 (74.80)	92.50 (74.17)	94.83 (77.03)	93.67 (75.60)
T ₂	93.67 (75.49)	95.33 (77.58)	94.50 (76.54)	94.00 (75.95)	94.00 (75.95)	94.00 (75.95)	93.83 (75.72)	94.67 (76.77)	94.25 (76.24)
T ₃	94.33 (76.47)	97.00 (80.12)	95.67 (78.30)	94.00 (75.95)	96.00 (78.72)	95.00 (77.33)	94.17 (76.21)	96.50 (79.42)	95.33 (77.82)
T ₄	97.67 (81.26)	98.33 (83.97)	98.00 (82.61)	96.00 (78.72)	98.00 (83.44)	97.00 (81.08)	96.83 (79.99)	98.17 (83.70)	97.50 (81.85)
T ₅	96.33 (79.14)	96.33 (79.14)	96.33 (79.14)	96.00 (78.72)	96.00 (78.72)	96.00 (78.72)	96.17 (78.93)	96.17 (78.93)	96.17 (78.93)
Mean	94.50 (76.78)	96.11 (79.13)		93.67 (75.77)	95.00 (77.74)		94.08 (76.28)	95.56 (78.43)	
	V	T	VxT	V	T	VxT	V	T	VxT
SEm (±)	0.57	0.99	1.40	0.73	1.27	1.80	0.44	0.76	1.07
CD @1%	2.28	3.96	NS	NS	5.07	NS	1.65	2.85	NS

NS- Non Significant

Figures in parenthesis are arcsine transformed values

Table 4: Effect of varieties, zinc oxide nanoparticle (ZnO NPs) and their interactions on Seedling length (cm) of cowpea.

Year	2023			2024			Pooled		
Treat/Var.	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
T ₀	21.80	22.20	22.00	23.30	24.20	23.75	22.55	23.20	22.88
T ₁	22.40	23.50	22.95	24.00	25.30	24.65	23.20	24.40	23.80
T ₂	22.10	25.30	23.70	24.40	25.60	25.00	23.25	25.45	24.35
T ₃	23.30	25.60	24.45	24.70	26.30	25.50	24.00	25.95	24.98
T ₄	25.60	28.60	27.10	25.90	29.50	27.70	25.75	29.05	27.40
T ₅	24.80	28.10	26.45	25.10	28.70	26.90	24.95	28.40	26.68
Mean	23.33	25.88		24.53	26.60		23.93	26.24	
	V	T	VxT	V	T	VxT	V	T	VxT
SEm (±)	0.25	0.43	0.60	0.27	0.48	0.67	0.18	0.31	0.43
CD @ 1 %	0.98	1.70	NS	1.09	1.90	NS	0.67	1.16	NS

NS- Non Significant

Table 5: Effect of varieties, zinc oxide nanoparticle (ZnO NPs) and their interactions on Seedling dry weight (g) of cowpea.

Year	2023			2024			Pooled		
Treat/Var.	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
T ₀	0.053	0.040	0.047	0.065	0.047	0.056	0.059	0.043	0.051
T ₁	0.055	0.041	0.048	0.066	0.049	0.057	0.061	0.045	0.053
T ₂	0.060	0.045	0.053	0.066	0.051	0.058	0.063	0.048	0.055
T ₃	0.062	0.048	0.055	0.067	0.052	0.059	0.064	0.050	0.057
T ₄	0.069	0.053	0.061	0.071	0.056	0.064	0.070	0.055	0.062
T ₅	0.066	0.051	0.059	0.069	0.053	0.061	0.067	0.052	0.060
Mean	0.061	0.046		0.067	0.051		0.064	0.049	
	V	T	VxT	V	T	VxT	V	T	VxT
SEm (±)	0.003	0.004	0.006	0.003	0.004	0.006	0.002	0.003	0.004
CD @ 1 %	0.011	NS	NS	0.011	NS	NS	0.007	NS	NS

NS- Non Significant

Table 6: Effect of varieties, zinc oxide nanoparticle (ZnO NPs) and their interactions on Vigour Index I of cowpea.

Year	2023			2024			Pooled		
Treat/Var.	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
T ₀	2006	2086	2046	2096	2225	2160	2051	2156	2103
T ₁	2083	2249	2166	2207	2377	2292	2145	2313	2229
T ₂	2069	2412	2240	2292	2405	2349	2181	2409	2295
T ₃	2196	2483	2340	2320	2523	2422	2258	2503	2381
T ₄	2500	2811	2656	2485	2890	2687	2493	2851	2672
T ₅	2388	2706	2547	2407	2754	2580	2398	2730	2564
Mean	2207	2458		2301	2529		2254	2493	
	V	T	VxT	V	T	VxT	V	T	VxT
SEm (±)	20.09	34.79	49.20	14.10	24.42	34.54	12.78	22.14	31.31
CD @ 1 %	80.07	138.68	NS	56.21	97.36	137.69	48.23	83.54	118.15

NS- Non Significant

Table 7: Effect of varieties, zinc oxide nanoparticle (ZnO NPs) and their interactions on Vigour Index II of cowpea.

Year	2023			2024			Pooled		
Treat/Var.	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
T ₀	4.87	3.76	4.32	5.89	4.31	5.10	5.38	4.04	4.71
T ₁	5.12	3.92	4.52	6.10	4.59	5.35	5.61	4.26	4.93
T ₂	5.63	4.29	4.96	6.20	4.82	5.51	5.92	4.55	5.24
T ₃	5.87	4.66	5.26	6.26	5.01	5.64	6.06	4.84	5.45
T ₄	6.74	5.22	5.98	6.86	5.50	6.18	6.80	5.36	6.08
T ₅	6.37	4.92	5.65	6.63	5.09	5.86	6.50	5.01	5.75
Mean	5.77	4.46		6.32	4.89		6.05	4.67	
	V	T	VxT	V	T	VxT	V	T	VxT
SEm (±)	0.25	0.43	0.60	0.26	0.45	0.64	0.16	0.28	0.40
CD @ 1 %	0.98	NS	NS	1.04	NS	NS	0.62	NS	NS

NS- Non Significant

Table 8: Effect of varieties, zinc oxide nanoparticle (ZnO NPs) and their interactions on Seed Mycoflora (%) of cowpea.

Year	2023			2024			Pooled		
Treat/Var.	V ₁	V ₂	Mean	V ₁	V ₂	Mean	V ₁	V ₂	Mean
T ₀	3.33 (6.14)	3.33 (6.14)	3.33 (6.14)	3.33 (6.14)	0.00 (0.00)	1.67 (3.07)	3.33 (6.14)	1.67 (3.07)	2.50 (4.61)
T ₁	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)
T ₂	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)
T ₃	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)
T ₄	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)
T ₅	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)	0.00 (0.00)
Mean	0.56	0.56		0.56	0.00		0.56	0.28	
	V	T	VxT	V	T	VxT	V	T	VxT
SEm (±)	0.98	1.69	2.39	0.72	1.25	1.77	0.61	1.05	1.49
CD @1%	NS	NS	NS	NS	NS	NS	NS	NS	NS

NS- Non Significant

Figures in parenthesis are arcsine transformed values

Conclusion

Variety Phule Sonali (V₁) had higher 100 seed weight, seedling dry weight, vigour index II while variety Phule Rakhumai (V₂) had higher germination percentage, shoot length, root length, seedling length, seedling vigour index I with lower electrical conductivity and seed mycoflora percentage. The foliar application of green synthesized ZnO NPs @ 125 ppm (T₄) had higher 100 seed weight, germination percentage, shoot length, root length, seedling length, seedling dry weight, vigour index I and vigour index II with the lower electrical conductivity and seed mycoflora percentage followed by ZnO NPs @ 150 ppm (T₅) which was superior over control. The interaction V₁T₄ (Phule Sonali + ZnO NPs @ 125 ppm) had higher 100 seed weight, seedling dry weight, vigour index II while the interaction V₂T₄ (Phule

Rakhumai + ZnO NPs @ 125 ppm) had higher germination percentage, shoot length, root length, seedling length, seedling vigour index I with lower electrical conductivity and seed mycoflora percentage during both the years on pooled basis.

References

- Abdul-Baki, A. A. and Anderson, J. D. (1973). Vigour determination multiple criteria. *Crop Sciences*, **13**, 630–633.
- Adhikari, T., Kundu, S., Biswas, A. K., Tarafdar, J. C. and Subba Rao, A. (2015). Characterization of zinc oxide nanoparticles and their effect on growth of maize (*Zea mays* L.) plant. *Journal of Plant Nutrition*, **38**(10), 1505–1515.
- Agarwal, V. K. (1976). Technique for the detection of seed borne fungi. *Seed Research*, **4**, 24–31.
- Alocilja, E. C. and Radke, S. M. (2003). Market analysis of biosensors for food safety. *Biosensors and Bioelectronics*,

- 18, 841–846. [https://doi.org/10.1016/S0956-5663\(03\)00049-3](https://doi.org/10.1016/S0956-5663(03)00049-3)
- Amin Farnia and Khodabandehloo, S. (2015). Changes in yield and its components of maize (*Zea mays* L.) to foliar application of zinc nutrient and mycorrhiza under water stress condition. *International Journal of Life Sciences*, **9**(5), 75–80.
- Anonymous. (1966). International Rules of Seed Testing. *Proceedings of the International Seed Testing Association*, **32**, 565–589.
- Arshi, N., Prashanthi, Y., Rao, T. N., Nageswara, N., Ahmed, F., Kumar, S. and Oves, M. (2020). Biosynthesis of ZnO nanostructures using *Azadirachta indica* leaf extract and their effect on seed germination and seedling growth of tomato: An eco-friendly approach. *Journal of Nanoelectronics and Optoelectronics*, **15**(11), 1412–1422.
- Arunachalam, P., Kannan, P., Prabukumar, G. and Govindaraj, M. (2013). Zinc deficiency in Indian soils with special focus to enrich zinc in peanut. *African Journal of Agricultural Research*, **8**(50), 6681–6688.
- Cobley, L. S. and Steele, W. M. (1976). *An introduction to the botany of tropical crops* (2nd ed.). Longman.
- Dhoke, S. K., Pramod, M., Rajashri, K. and Anand, K. (2013). Effect of nanoparticles suspension on the growth of mung (*Vigna radiata*) seedlings by foliar spray method. *Nanotech. Development*, **3**(1), 1–5.
- Dobariya, V. A. and Patel, A. P. (2021). Effect of iron and zinc on growth, yield and quality of cowpea (*Vigna unguiculata* L.) under South Gujarat condition. *International Journal of Current Microbiology and Applied Sciences*, **10**(11), 346–351.
- Gayathri, S. (2019). *Seed invigoration with inorganic nanoparticles in chillies (Capsicum annuum L.)* (M.Sc. thesis). Kerala Agricultural University, Thrissur.
- Gokak, I. B. and Taranath, T. C. (2015). Morphological and biochemical responses of *Abelmoschus esculentus* (L.) Moench to zinc nanoparticles. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, **6**(2), 025017.
- Hanumanthappa, D. C., Sushmitha, B. P. and Ganesh, A. S. (2019). Standardization of nano boron and nano zinc concentrations for effective cultivation of groundnut (*Arachis hypogaea* L.). *International Journal of Chemical Studies*, **7**(3), 2720–2723.
- ISTA. (2013). International Rules of Seed Testing. *Seed Science & Technology*, **27**, 25–30.
- Kumar, S. M., Yadav, S. K., Yadav, S., Choudhary, R., Lal, S. K., Bharadwaj, C., Hussain, Z. and Kumar, P. R. (2023). Preliminary studies on the effect of nanoparticle seed treatments on seed quality attributes and yield in chickpea. *Turkish Journal of Agriculture and Forestry*, **47**, 100–115.
- Lakshmi, S. J., Roopabai, R. S., Sharanagouda, H., Ramachandra, C. T., Sushila, N. and Shivanagouda, R. D. (2017). Biosynthesis and characterization of ZnO nanoparticles from spinach (*Spinacia oleracea*) leaves and its effect on seed quality parameters of green gram (*Vigna radiata*). *International Journal of Current Microbiology and Applied Sciences*, **6**(9), 3376–3384.
- Laware, S. L. and Raskar, S. V. (2014). Influence of zinc oxide nanoparticles on growth, flowering and seed productivity in onion. *International Journal of Current Microbiology and Applied Sciences*, **3**(7), 874–881.
- Maithreyee, M. N. and Gowda, R. (2015). Influence of nanoparticles in enhancing seed quality of aged seeds. *Mysore Journal of Agricultural Sciences*, **49**, 310–313.
- Mendez-Arguello, B., Vera-Reyes, I., Mendoza-Mendoza, E., García-Cerda, L. A., Puente-Urbina, B. A. and Lira-Saldivar, R. H. (2016). Growth promotion of *Capsicum annuum* plants by zinc oxide nanoparticles. *Nova Science*, **8**, 140–156.
- Miralles, P., Church, T. L. and Harris, A. T. (2012). Toxicity, uptake, and translocation of engineered nanomaterials in vascular plants. *Environmental Science & Technology*, **46**(17), 9224–9239.
- Nalini, P., Gupta, B. and Pathak, G. C. (2013). Foliar application of Zn at flowering stage improves plants performance, yield and yield attributes of blackgram. *Indian Journal of Experimental Biology*, **51**, 548–555.
- Obata, H., Kawamura, S., Senoo, K. and Tanaka, A. (1999). Changes in the level of protein and activity of Cu/Zn-superoxide dismutase in zinc deficient rice plant, *Oryza sativa* L. *Soil Science and Plant Nutrition*, **45**, 891–896.
- Pandey, N., Pathak, G. C. and Sharma, C. P. (2006). Zinc is critically required for pollen function and fertilisation in lentil. *Journal of Trace Elements in Medicine and Biology*, **20**, 89–96.
- Pandey, A. C., Sanjay, S. S. and Yadav, S. R. (2010). Application of ZnO nanoparticles in influencing the growth rate of *Cicer arietinum*. *Journal of Experimental Nanoscience*, **5**(6), 488–497.
- Patra, P., Choudhury, S., Mandal, S., Basu, A., Goswami, A., Gogoi, R., Srivastava, C., Kumar, R. and Gopal, M. (2013). Effect of sulfur and ZnO nanoparticles on stress physiology and plant (*Vigna radiata*) nutrition. In *Advanced Nanomaterials and Nanotechnology* (Springer Proceedings in Physics, Vol. **143**). Springer.
- Peiman, M. R., Zahra, M., Mehdi, G. and Majid, R. (2022). Effects of foliar application of zinc oxide nanoparticles on traits of several medicinal plants under aeroponic system conditions. *International Journal of Horticultural Science and Technology*, **9**(4), 445–452.
- Prasad, T. N., Sudhakar, V. P., Sreenivasulu, Y., Latha, P., Munaswamy, V., Raja, R. K., Sreeprasad, T. S., Sajanlal, P. R. and Pradeep, T. (2012). Effect of nanoscale zinc oxide particles on the germination, growth and yield of peanut. *Journal of Plant Nutrition*, **35**, 905–927.
- Presley, J. T. (1958). Relations of protoplast permeability of cotton seed viability and pre deposition of disease. *Plant Disease Research*, **42**, 582.
- Purseglove, J. W. (1976). *Tropical crops: Dicots*. Longman Group Ltd.
- Rai-Kalal, P. and Jajoo, A. (2021). Priming with zinc oxide nanoparticles improve germination and photosynthetic performance in wheat. *Plant Physiology and Biochemistry*, **160**, 341–351.
- Rawat, P. S., Kumar, R., Ram, P. and Pandey, P. (2018). Effect of nanoparticles on wheat seed germination and seedling growth. *International Journal of Agricultural and Biosystems Engineering*, **12**(1), 13–16.
- Sedghi, M., Hadi, M. and Toluie, S. G. (2013). Effect of nano zinc oxide on the germination of soybean seeds under drought stress. *Annales of West University of Timisoara. Series Biology*, **2**, 73–78.
- Sellschop, J. P. F. (1962). Cowpea (*Vigna unguiculata* L. Walp). *Field Crops Abstracts*, **15**, 259–266.

- Dhoke, S. K., Mahajan, P., Kamble, R. and Khanna. (2013). Effect of nanoparticles suspension on the growth of mung (*Vigna radiata*) seedlings by foliar spray method. *Nanotech Development*, **3**, 1–5.
- Snehalbhai, P. (2016). *Effect of ZnO nanoparticles on germination, growth and yield of groundnut (Arachis hypogaea L.)* (Ph.D. Thesis). Anand Agricultural University, Anand, Gujarat.
- Sturikova, H., Krystofova, O., Huska, D. and Adam, V. (2018). Zinc, zinc nanoparticles and plants. *Journal of Hazardous Materials*, **349**, 101–110.
- Tarafdar, J. C., Agarwal, A., Raliya, R., Kumar, P., Burman, U. and Kaul, R. K. (2013). ZnO nanoparticles induced synthesis of polysaccharides and phosphates by *Aspergillus* fungi. *Advanced Science, Engineering and Medicine*, **4**, 1–5.
- Tejaswini, K. S., Kurnalliker, V. K., Shakuntala, N. M., Macha, S. I., Khan, H. and Hiregoudar, S. (2019). Studies on efficacy of nanoparticles in improving seed physiological parameters in groundnut (*Arachis hypogaea* L.). *International Journal of Chemical Studies*, **7**(5), 1786–1791.
- Tondey, M., Kalia, A., Singh, A., Abd Elsalam, K., Hassan, M. and Dher, G. (2022). A comparative evaluation of the effects of seed invigoration treatments with precursor zinc salt and nano-sized zinc oxide (ZnO) particles on vegetative growth, grain yield and quality characteristics of *Zea mays*. *Journal of Analytical Science and Technology*, **13**, 40.
- Valadkhan, M., Mohammadi, K. and Nezhad, M. T. K. (2015). Effect of priming and foliar application of nanoparticles on agronomic traits of chickpea. *Biological Forum*, **7**(2), 599–602.
- Velasco, P. E. A., Betancourt Galindo, R., Valdez Aguilar, L. A., Gonzalez Fuentes, J. A., Puente Urbina, B. A., Lozano Morales, S. A. and Sánchez Valdés, S. (2020). Effect of the morphology, surface modification and application methods of ZnO-NPs on the growth and biomass of tomato plants. *Molecules*, **25**(6), 1282.
- Wankhade, N. J. and Shelar, V. R. (2025). Impact of biosynthesized zinc oxide nanoparticles (ZnO NPs) and silver nanoparticles (Ag NPs) on seed production of onion (*Allium cepa* L.). *Plant Archives*, **25**(2), 1235–1248.
- Zhang, F., Wang, R., Xiao, Q., Wang, Y. and Zhang, J. (2006). Effects of slow/controlled release fertilizer cemented and coated by nano-materials on biology. II. Effects of slow/controlled-release fertilizer cemented and coated by nano-materials on plants. *Nanoscience*, **11**, 18–26.
- Zulfiqar, U., Hussain, S., Ishfaq, M., Matloob, A., Ali, N., Ahmad, M., Alyemeni, M. N. and Ahmad, P. (2020). Zinc-induced effects on productivity, zinc use efficiency, and grain biofortification of bread wheat under different tillage permutation. *Agronomy*, **10**(2). <https://doi.org/10.3390/agronomy10101566>